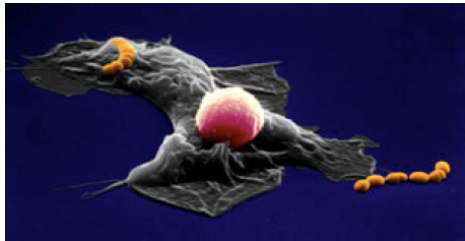
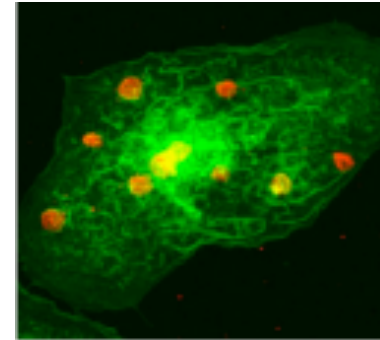


How one cell eats another –

Experiments and modelling elucidate early signalling events and biophysical requirements for uptake



Robert Endres

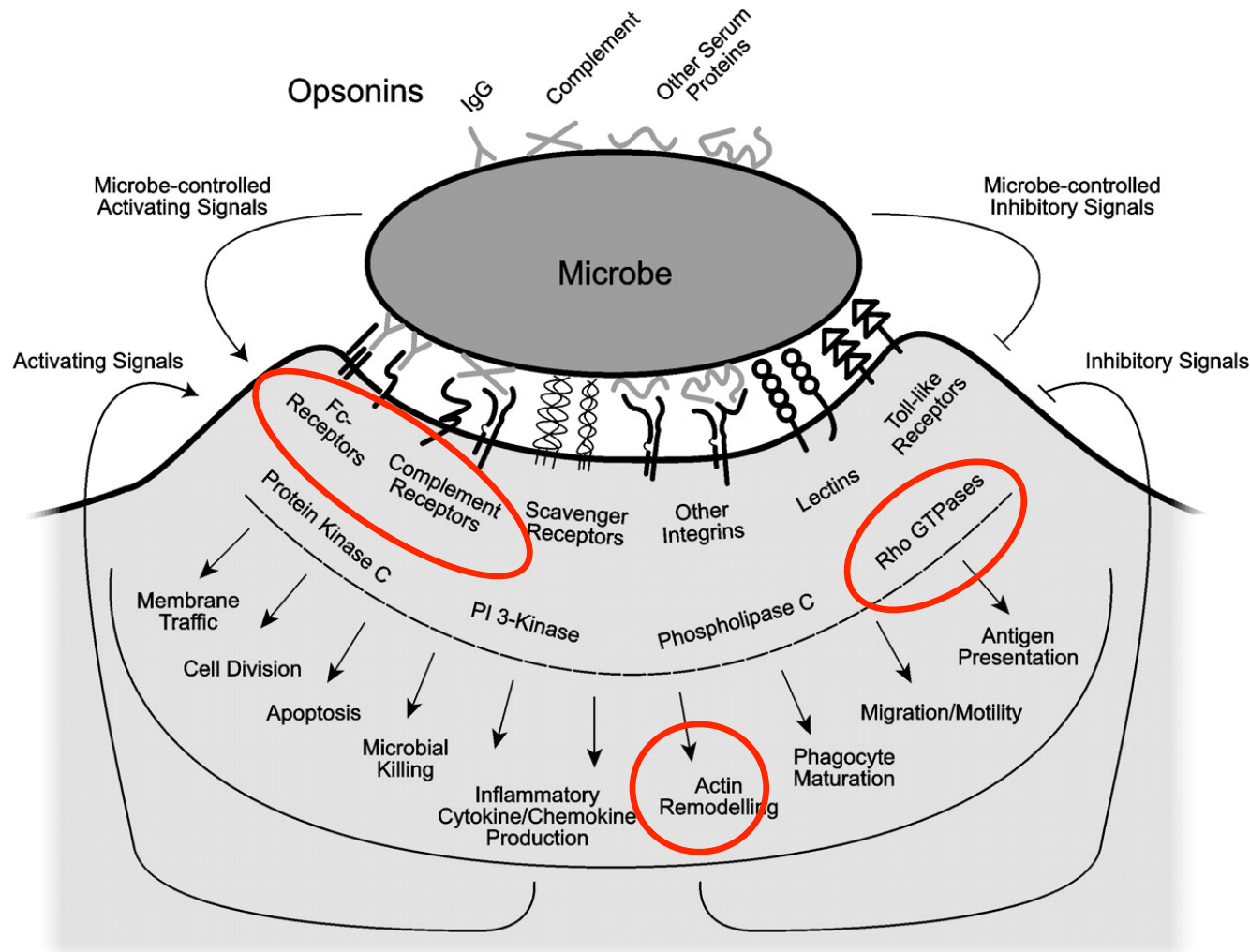


Biological Physics Group at Imperial College:

<http://www3.imperial.ac.uk/biologicalphysics>

IOP – Complexity in Biological Systems – Bath, 20 May 2010

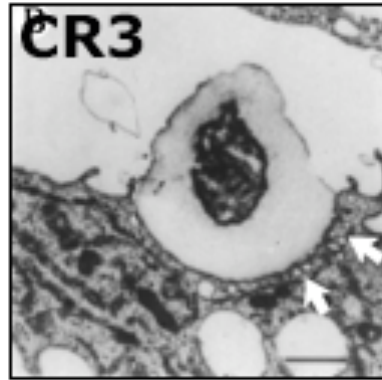
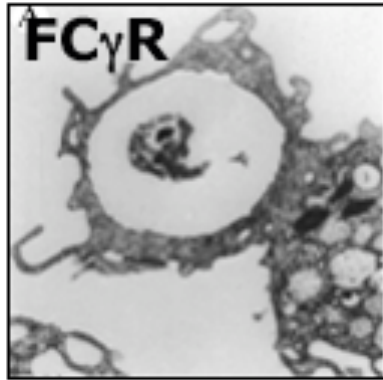
Daunting signalling complexity in phagocytosis



about 140
different molecular
species are involved

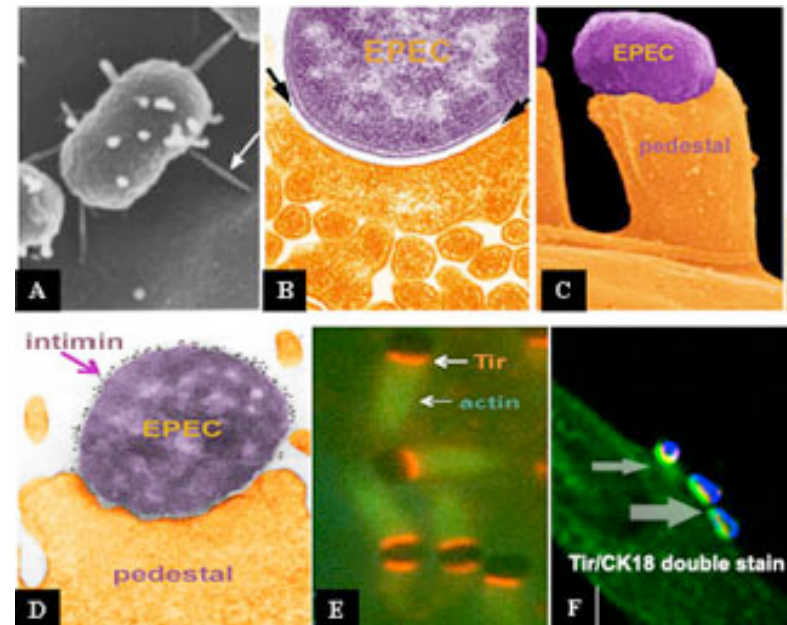
Underhill & Ozinsky (2002)

Actin-rich protrusions in phagocytosis...



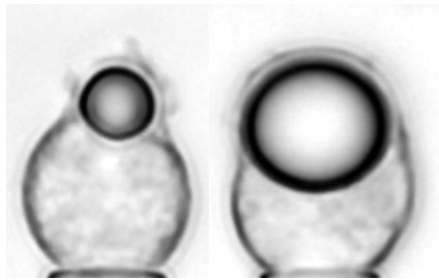
Allen & Aderem (1996)

...and anti-phagocytosis



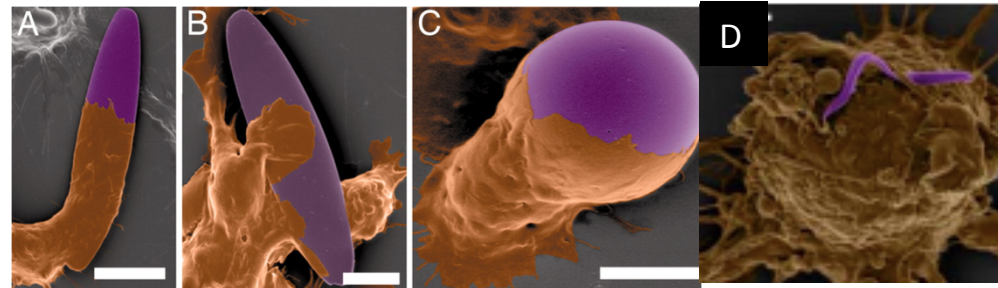
Universal biophysical aspects of phagocytosis

1. Size (in)dependence (endocytosis vs phagocytosis)



Herant et al. (2006)

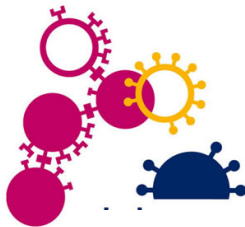
2. Shape dependence



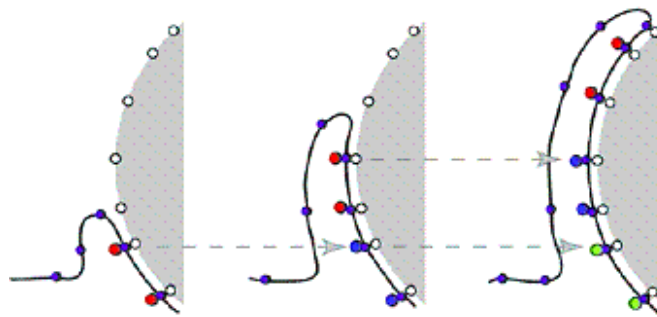
Champion et al. (2006,2009)

3. Elastic properties

4. Ligand density →



Zipper mechanism



Griffin et al. (1975), Swanson (2008)

Outline

1. Minimal biophysical model of early uptake in phagocytosis

Cup-shape and energetic requirements

2. Experimental verification using Fcy-R and IgG-coated latex particles

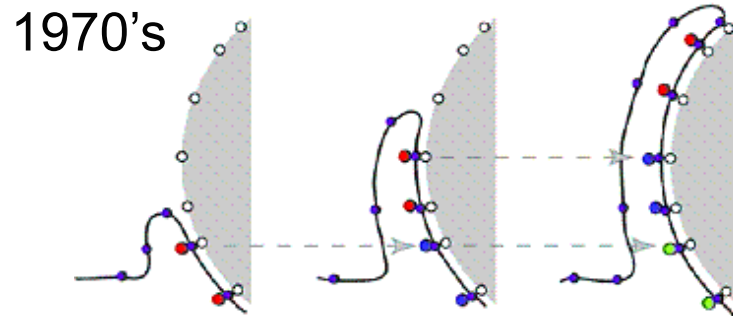
3. Work in progress - extension of model and new experiments

Model of acto-myosin cytoskeleton

Experimental role of myosin isoforms

4. Summary

The Zipper mechanism: generally accepted but untested

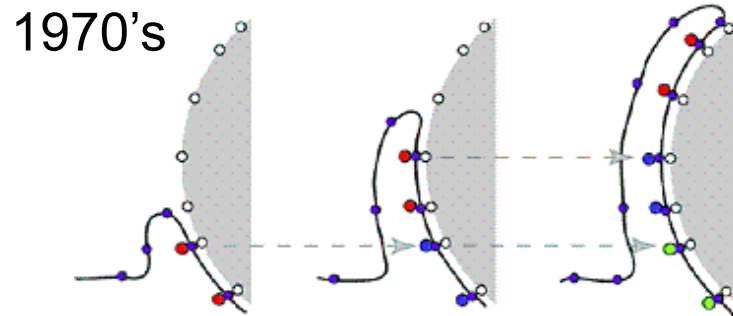


Zipper mechanism:

Unidirectional, sequential ligand
-receptor interactions guide membrane
around particle.

→ Can it explain biophysical aspects?

The Zipper mechanism: generally accepted but untested

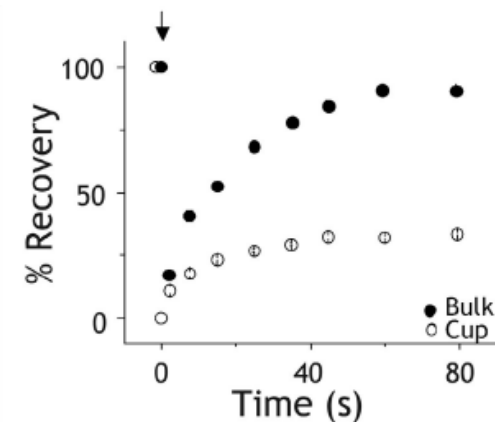
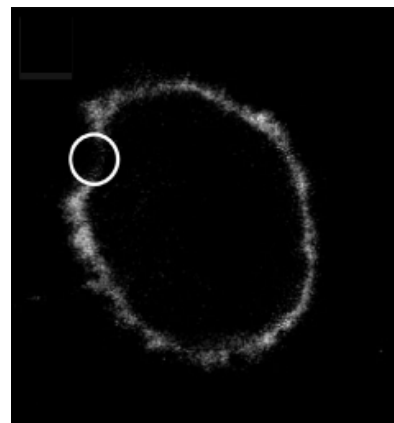
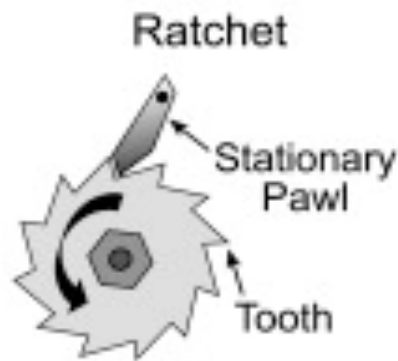


Zipper mechanism:

Unidirectional, sequential ligand-receptor interactions guide membrane around particle.

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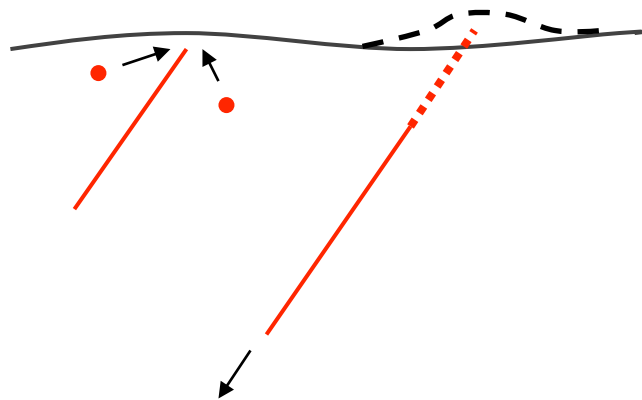
Ratchet-like mechanism ? FRAP: Immobilization of proteins and lipids in cups



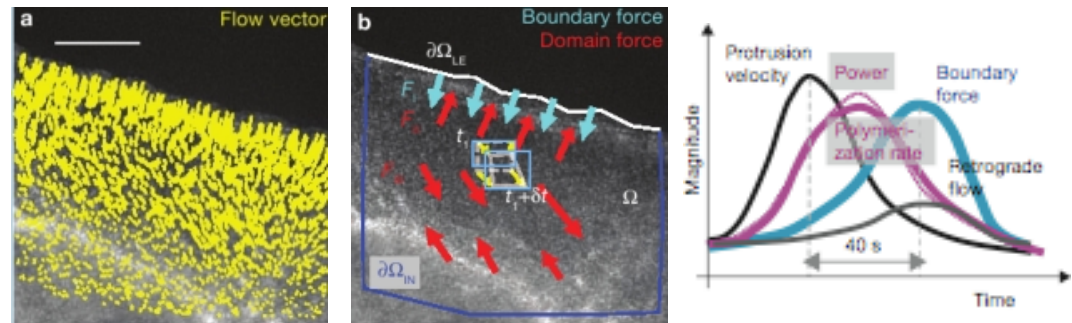
Corbett-Nelson et al. (2006)

Model ingredients

Actin polymerizes at barbed end

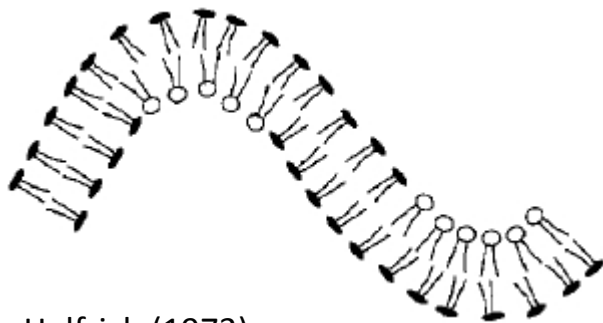


Fluorescent speckle microscopy



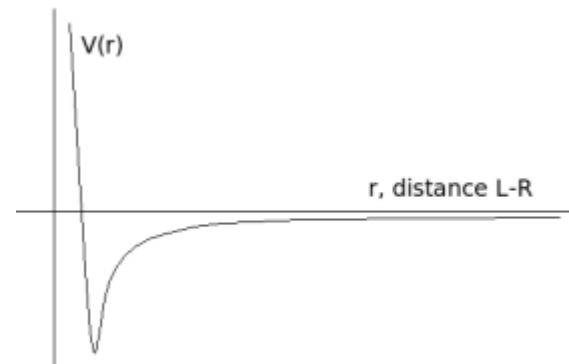
Ji et al. (2008)

Membrane energy: bending, surface tension, volume constraint

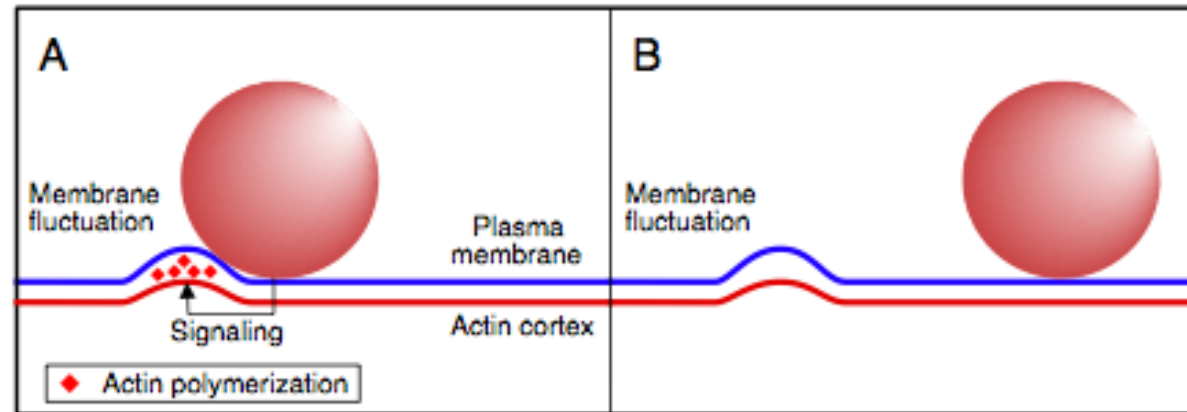


Helfrich (1973)

Ligand-receptor binding



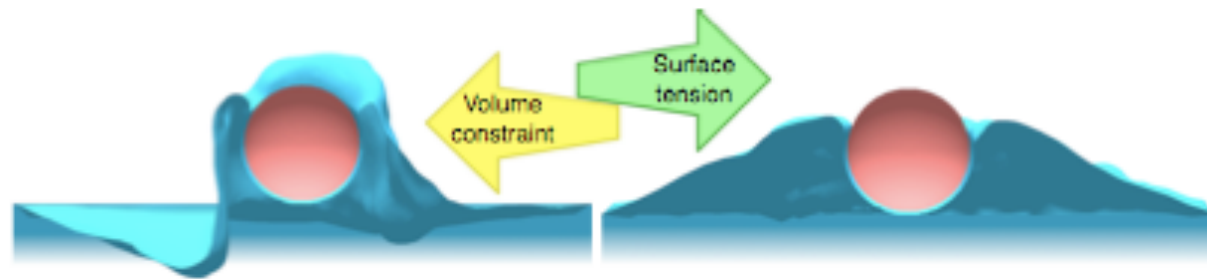
Minimal biophysical model for zipper mechanism



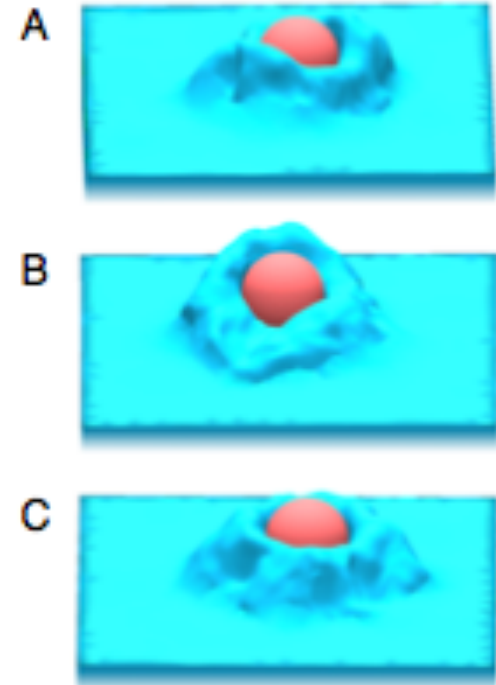
- (1) Random, thermal membrane fluctuation.
- (2a) If near particle, ligand-receptor binding leads to actin polymerization, stabilizing fluctuation \rightarrow irreversible \rightarrow ratchet.
- (2b) If away from particle, no stabilization and membrane fluctuation may be reversed at a later time.
- (3) Model implemented with finite-element simulations and Monte Carlo algorithm.



Engulfment over wide range of parameters



different parameters
→ robustness

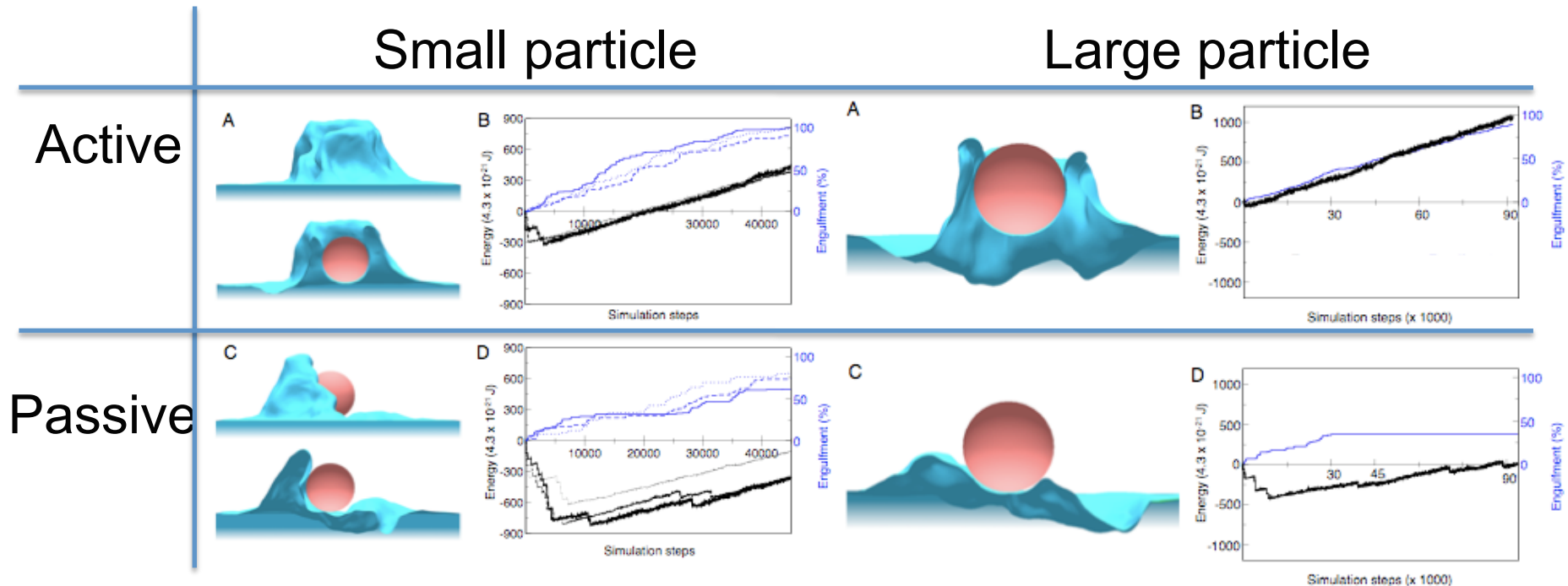


same parameters
→ variability

How important is actin-polymerization?

Would “passive” ligand-receptor binding be sufficient?

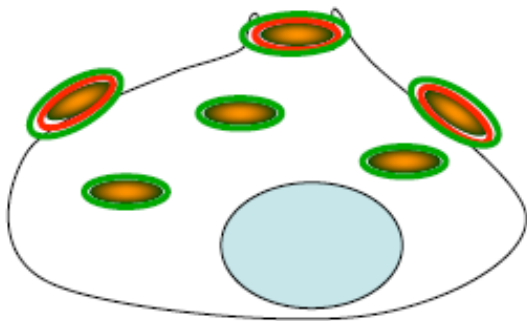
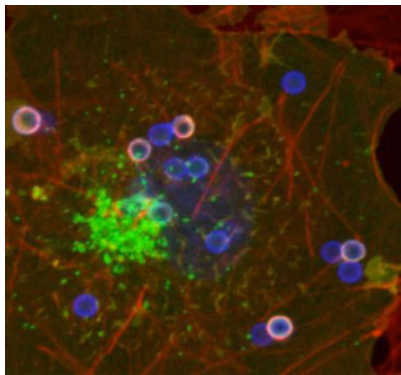
“Active” versus “passive” engulfment



- Passive engulfment still works for small particles, but is much slower and cups are more variable.
- Can we experimentally test these two predictions?

Imaging of phagocytosis using transfected fibroblasts

Phagocytic assay



Time series data of FcR dynamics during uptake (imaging of 3 μ m particles)

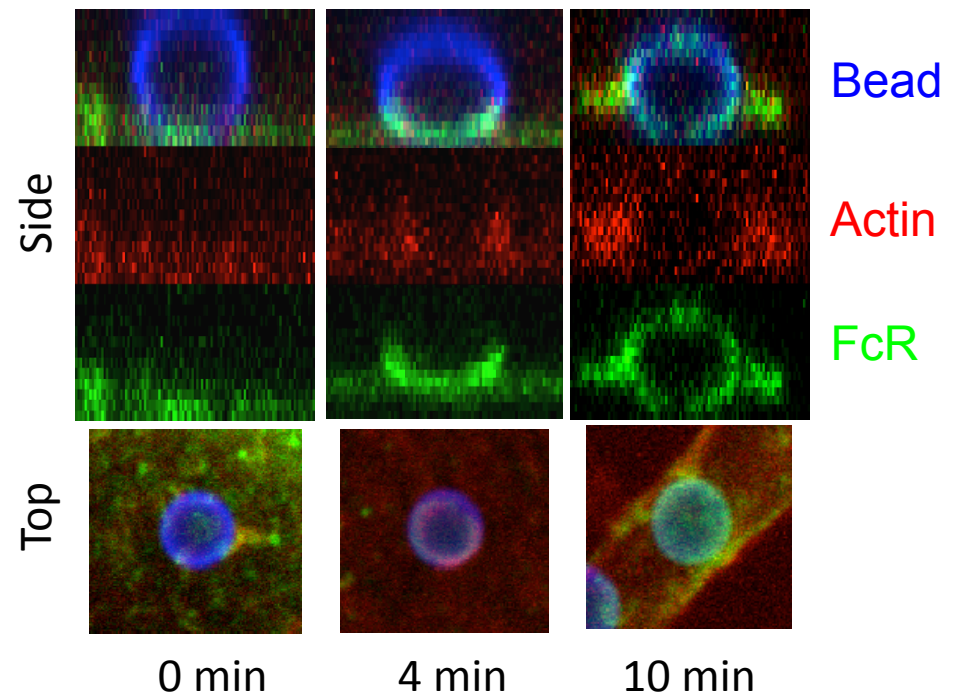
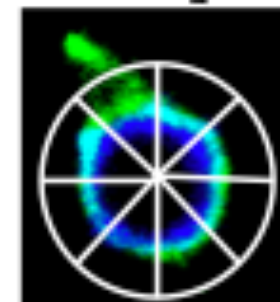
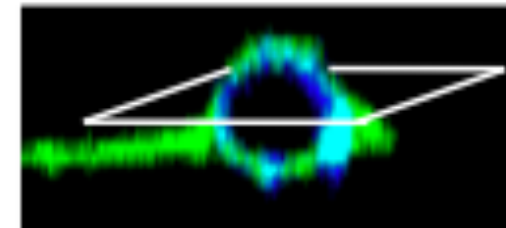
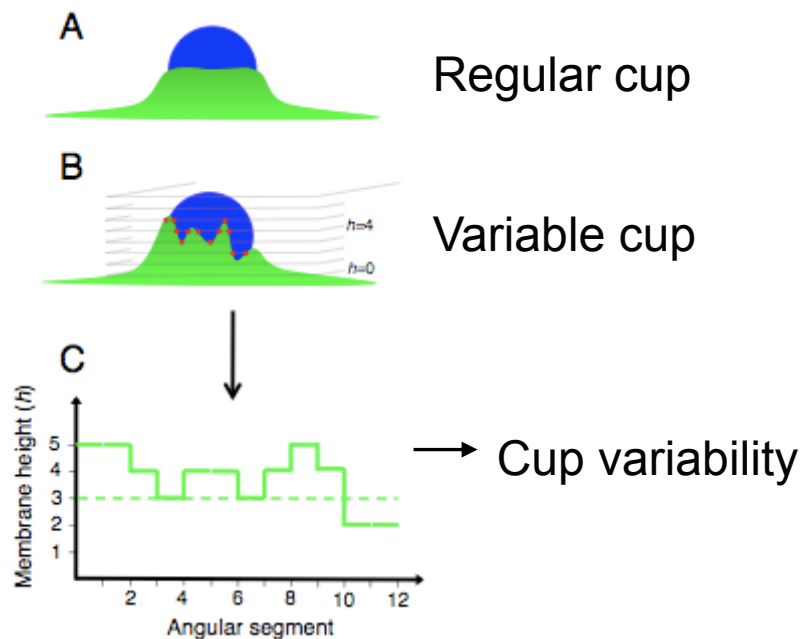


Image analysis of 3d-data from confocal microscopy

(a) Cells expressing wild-type Fc receptor = **active zipper**

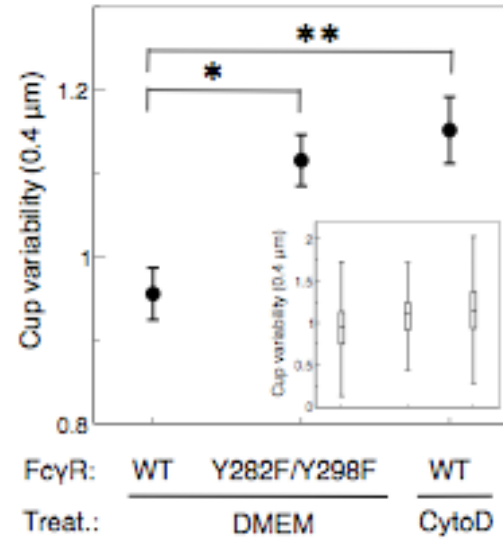
(b) Cells expressing signalling-dead mutant receptor } **passive**

(c) Cells transfected with cytochalasin D } **zippers**



Experiments confirm our predictions

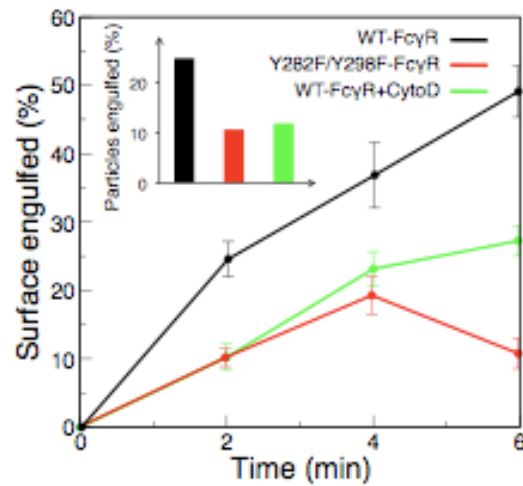
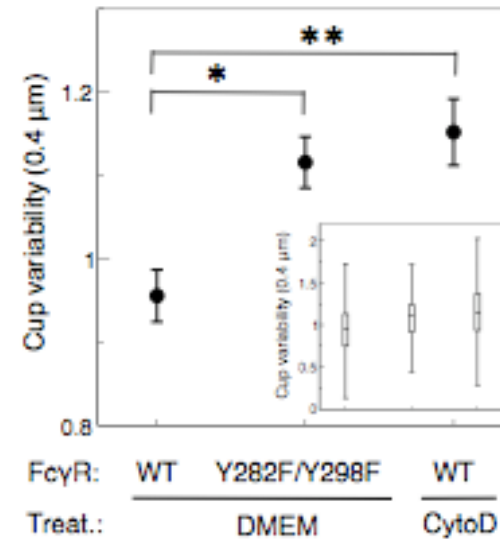
Passive-zipper cells
have more variable cups



Experiments confirm our predictions

Passive-zipper cells
have more variable cups

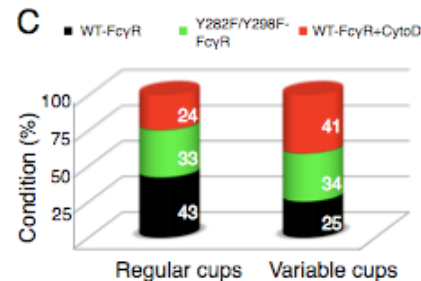
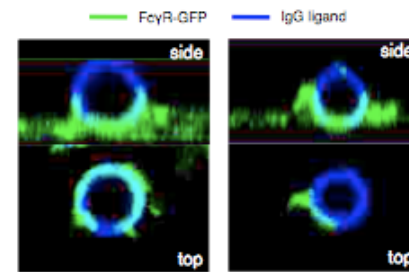
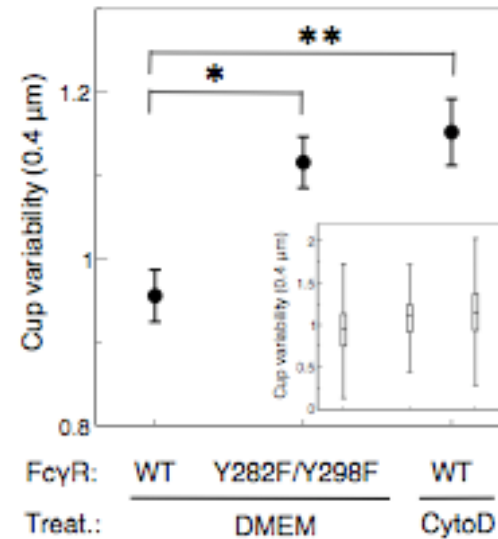
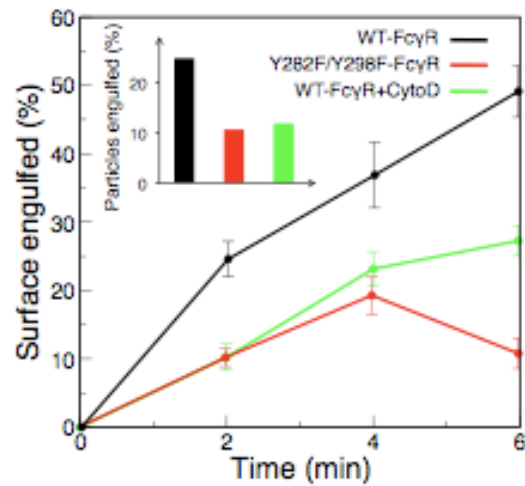
Passive-zipper cells
engulf more slowly



Experiments confirm our predictions

Passive-zipper cells
have more variable cups

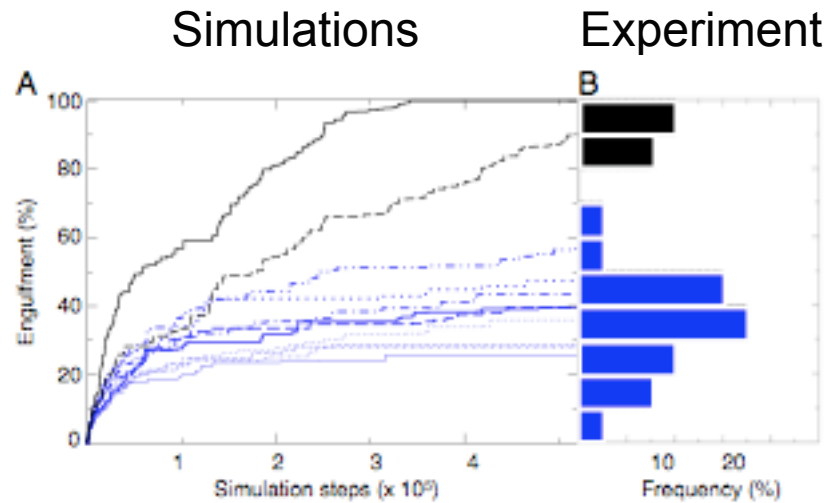
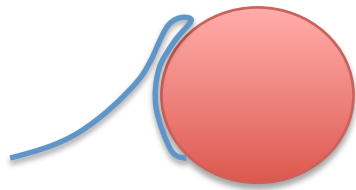
Passive-zipper cells
engulf more slowly



Universal
features in
cup shape

Model confirms mechanical bottleneck....

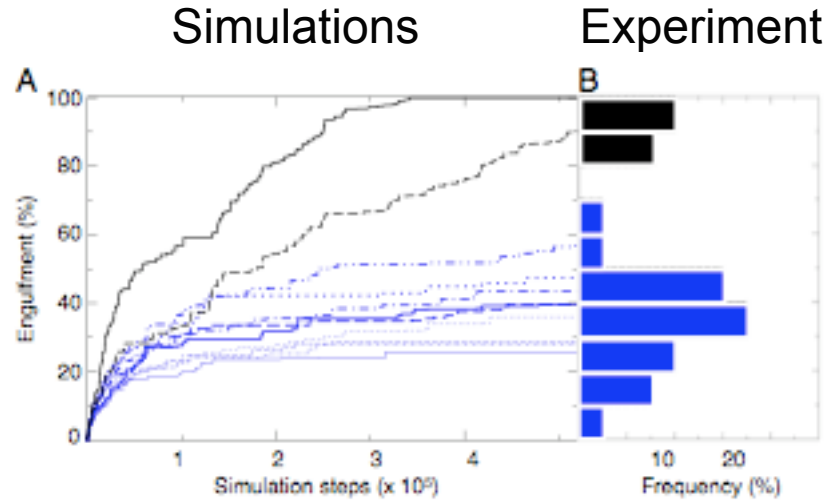
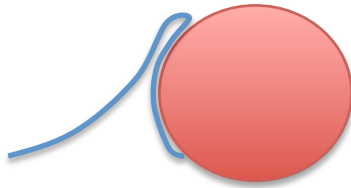
Mechanical bottleneck



Bimodal distribution from experiments with WT-FcR after 10 min.

Model confirms mechanical bottleneck....

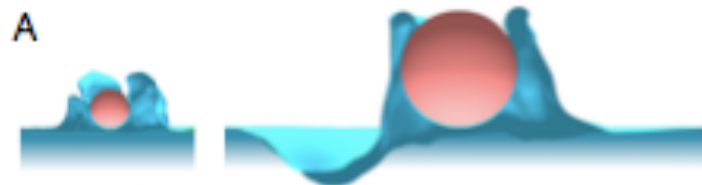
Mechanical bottleneck



Bimodal distribution from experiments with WT-FcR after 10 min.

....and shape dependence

Size independence

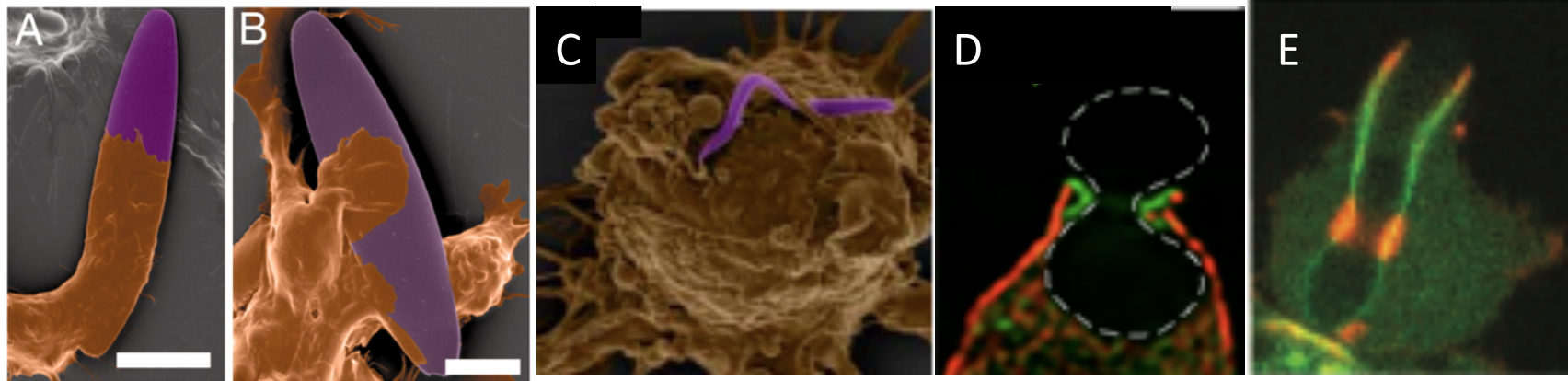


Shape dependence



→ Model difficulties with cup closure

New improved model:

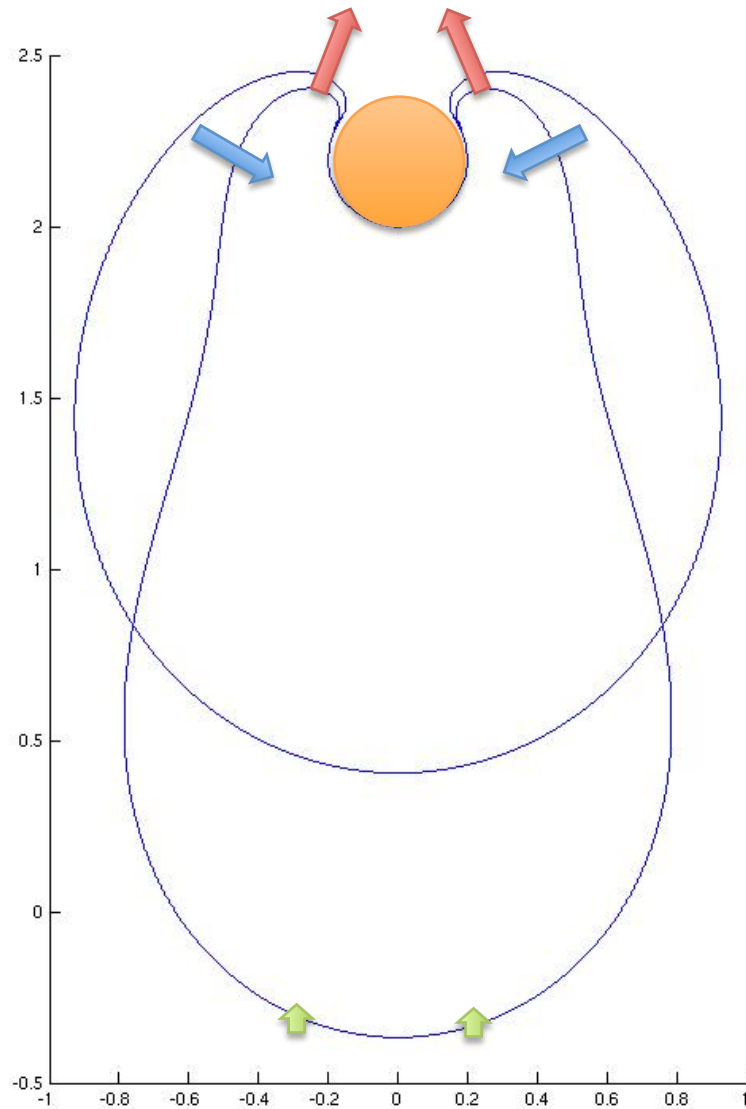


Address the following fundamental questions:

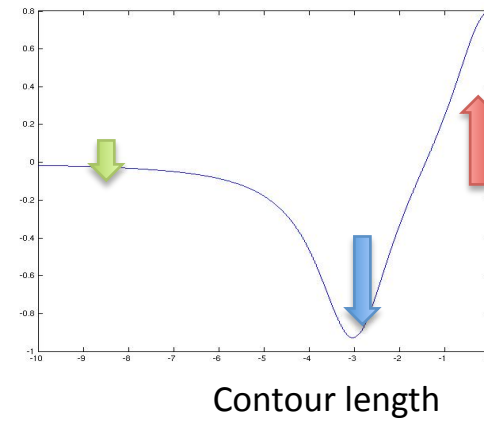
- (1) Particle-shape dependence
- (2) Cup closure, particle squeezing
- (3) Actin waves at cup
- (4) Particle stiffness dependence

→ Extend model to include acto-myosin cytoskeleton

Preliminary: cell shape for given cell-pressure profile



Applied pressure profile:



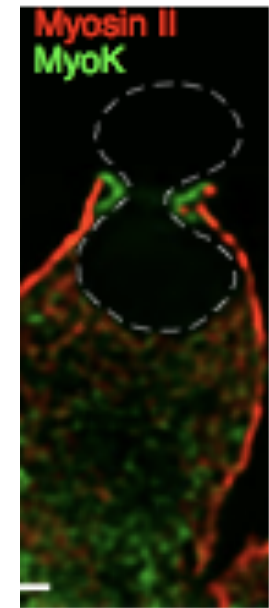
Next: calculate pressure from model for acto-myosin cytoskeleton and signalling

New experiments:

1) Role of motor proteins in cup closure and particle squeezing

Imaging by confocal microscopy of motor proteins using anti-bodies and GFP-constructs

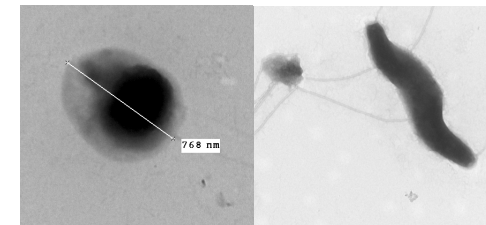
Knock-downs by RNAi and imaging by SEM



2) Shape dependence of bacterial uptake

Human pathogen *Campylobacter*

Unfortunately very small, taken up by epithelia via endocytosis, avoids lysosome

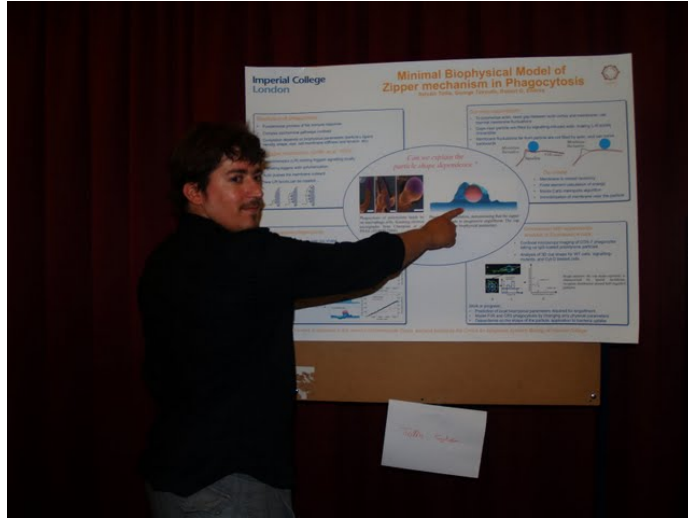


Coccoid and helical shapes

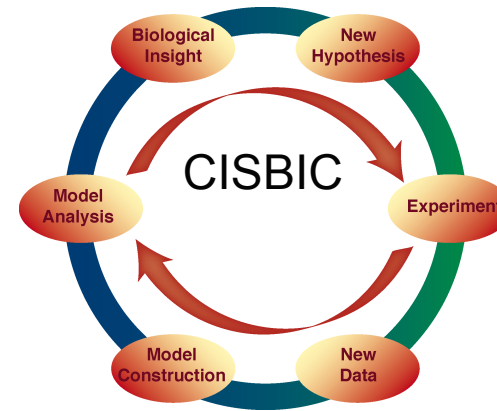
Summary

- Importance of **universal biophysical requirements** in phagocytosis:
(a) ligand density, (b) particle shape and (c) stiffness.
- **Minimal biophysical model** uses ratchet-like mechanism for zipper.
- **Model predictions** for active and passive zippers:
Removal of actin-driven processes leads to slower engulfment and highly variable cups. This is confirmed by experiments.
- **Active zipper:** shape but not size matters.
Passive zipper: size matters as well.
- Active processes make phagocytosis **robust**, and enable engulfment of large particles.
- **New developments:** model of acto-myosin cytoskeleton and experiments and myosins.

Acknowledgements



Sylvain Tollis in Croatia '09



- Members of the Biological Physics group:
 - Sylvain Tollis
 - Anna Dart
 - George Tzircotis

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"Engineering Principles in Chemotaxis
Signalling Pathways"