# **Setup for Live Imaging using the WF1 Microscope**

# Please ask for help if you are unsure

| General considerations               | 1 |
|--------------------------------------|---|
| Heating                              | 2 |
| Turning on                           | 2 |
| To adjust the temperature:           | 2 |
| Humidity                             | 2 |
| CO <sub>2</sub>                      | 2 |
| Lid options                          | 3 |
| Turning on                           | 4 |
| In-Line safety clips                 | 4 |
| Gas Cylinder Safety Check            | 4 |
| Turn on CO <sub>2</sub> gas cylinder | 5 |
| Setting flow rate                    | 5 |
| Shutting down                        | 6 |
| CO <sub>2</sub>                      | 6 |
| Heating                              |   |

## **General considerations**

#### Choice of Media

 Phenol Red dye in culture media has low-level auto-fluorescence particularly in the green emission wavelengths, which can interfere with the fluorescent signal, and is best avoided. There are Phenol Red free media available for fluorescent imaging.

#### Well Liquid levels

Imaging for several hours or more at 37°C can cause evaporation of the
culture/imaging media. Make sure each well has sufficient liquid level to
compensate for this. Also filling unused wells in the same plate can be
beneficial. Placing a container of water at the base of the microscope
incubation chamber helps to provide a humid environment (an old pipette box
with about 1cm of water is usually sufficient).

# **Heating**

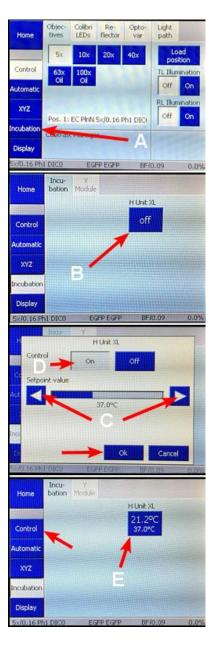
The microscope incubator chamber can be set to a specified temperature. Turn on at least 30 minutes before use to allow the chamber to reach temperature and equilibrate. For longer term imaging (time series) then preheat the chamber for at least 2 hours before use, to prevent focus drift).

## Turning on

- Make sure all incubator doors are closed.
- Switch on the plug on the wall marked heating (NB the Heating unit needs to be switched on before the "Main" microscope controls otherwise it is not recognised by the system – in this case switch off the Main switch and then back on again)
- On the TFT screen, select "Microscope" and then "Inkubation" (A) (if Inkubation is not visible then restart the system as above).
- The display will show the temperature of the H unit XL as OFF

## To adjust the temperature:

- Press the OFF button (B) to open the temperature controls. Select the required temperature (C) and then press ON (D). Then press OK.
- The display will now show the current and the required temperatures of the incubator (E).
- Press the Control button to return to the objective display



# **Humidity**

Imaging of live samples will result in evaporation of the growth media supporting the cells. This will become a significant problem for longer term imaging at higher temperatures (eg 37<sub>0</sub>C). This evaporation can be significantly reduced by placing a container (a base of a used pipette tip box is ideal) partially filled (1cm deep) with water inside the base of the microscope incubation chamber but away from the stage movement. If running over several days this box should be checked and filled daily. Take care not to spill water inside the microscope incubator chamber

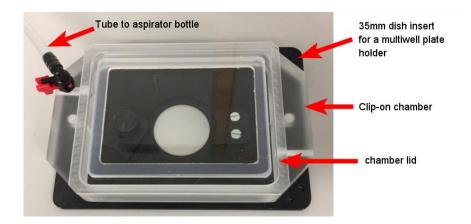
# $CO_2$

If using CO<sub>2</sub> please check cylinder pressure a few days before required in case the cylinder needs replacing – see checking method below

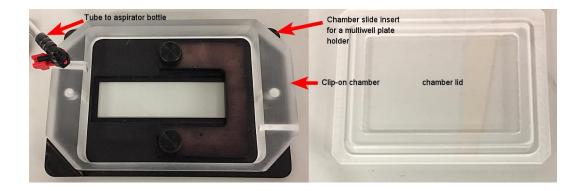
Please note that the microscope incubator chamber, as a whole unit, cannot maintain a controlled CO<sub>2</sub> environment. Instead, it is possible to supply a 5% CO<sub>2</sub>/air supply to a specific sample carrier lid. There are several lid options for 35mm dishes, chamber slides and multi-well plates. Each is supplied with tubing to connect to the 5% gas supply

# Lid options

• 35mm dish



Chamber slide



• Multi-well plate



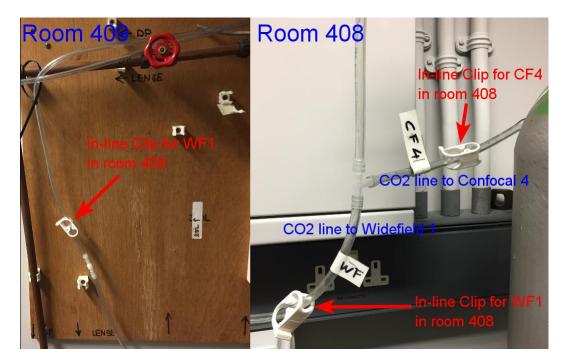
### **Turning on**

- Check the level of distilled water in the humidifying aspirator bottles inside the incubator chamber should be about a cm full in each (b)
- Place the lid you are using over your sample and connect to the aspirator bottle – black connector (a)



NB. The gas cylinder for the WF1 is in Room 408 and is shared between the WF1 and the CF4. There are 3 in-line safety clips to check.

# In-Line safety clips

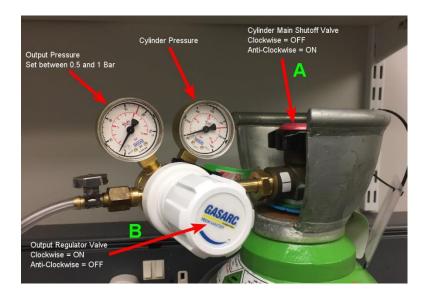


- Room 409 open the in-line clip on the far wall
- Room 408 open the clip to the WF2 and check the clip to CF4 is closed unless it is already in use

# **Gas Cylinder Safety Check**

If **not** already in use on the confocal 4:

- Check gauges: (there may be some residual pressure from the last use)
  - o left had gauge should be between 0 and 1 BAR
  - right hand gauge should be between 0 and last used cylinder pressure value
- Check the main cylinder valve is closed fully clockwise (A)
- Check the pressure regulator valve is closed (B) it should feel loose (almost fully anticlockwise) – it will feel stuck if turned too far anticlockwise
- Check visually the pipework form cylinder to CO<sub>2</sub> controller



## Turn on CO<sub>2</sub> gas cylinder

(NB if already being used by the CF4, just check and adjust pressure regulator valve (B) as necessary)

- Open the main cylinder valve (A) about I turn (anticlockwise) a positive pressure value should be displayed in the right-hand gauge showing the gas level in the cylinder (open more if no pressure showing). If pressure displayed is still zero, then the cylinder may be empty - contact FILM staff
- Then by turning the pressure regulator valve (B) slowly clockwise, increase
  the pressure to between 0.5 and 1 BAR as displayed in the left hand (output)
  gauge a slight resistance will be felt when the valve starts to open.

#### **Setting flow rate**

- Switch on the plug on the wall marked CO<sub>2</sub>
- Turn on CO<sub>2</sub> controller to the right of the microscope (a)
- Check for bubbling inside the aspirator bottles inside the incubator
- If nothing is immediately visible, press and hold the purge button (b) on the front of the CO2 controller until bubbling appears.
   Release when bubbles appear.



- The flow (bubble) rate can be set (approximately 2-5 bubbles per second) by turning the flow control on the front of the CO2 unit (c)
- Re-Check the regulator output gauge (B) again and adjust the regulator valve as necessary to maintain 0.5 to 1 BAR

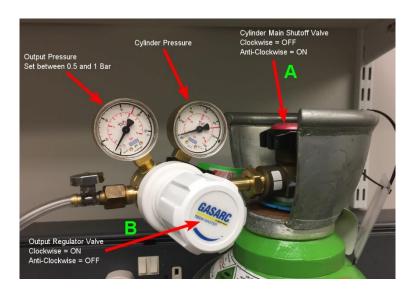
# **Shutting down**

#### 

- Turn off CO<sub>2</sub> controller to the right of the microscope and turn off at the wall
- Close the in-line safety clips (rooms 408 and 409)

### If not being used on the CF4:

- Check the main cylinder valve (A) is closed fully clockwise
- Check the pressure regulator valve (B) is closed almost fully anticlockwise it should feel loose – it will feel stuck if turned too far anticlockwise



### **Heating**

- Switch off the temperature on the TFT control unit (incubation) as described above and turn off at the wall
- Remove humidifier box if used.